

**Phase behavior of two-component lipid membranes: Theory and experiments**Md. Arif Kamal,<sup>1</sup> Antara Pal,<sup>1</sup> V. A. Raghunathan,<sup>1</sup> and Madan Rao<sup>1,2</sup><sup>1</sup>*Raman Research Institute, C. V. Raman Avenue, Bangalore 560 080, India*<sup>2</sup>*National Centre for Biological Sciences, Tata Institute of Fundamental Research, Gandhi Krishi Vignyan Kendra Campus, Bangalore 560 065, India*

(Received 1 February 2012; published 18 May 2012)

The structure of the ripple phase of phospholipid membranes remains poorly understood in spite of a large number of theoretical studies, with many experimentally established structural features of this phase unaccounted for. In this article we present a phenomenological theory of phase transitions in single- and two-component achiral lipid membranes in terms of two coupled order parameters: a scalar order parameter describing lipid chain melting, and a vector order parameter describing the tilt of the hydrocarbon chains below the chain-melting transition. This model reproduces all the salient structural features of the ripple phase, providing a unified description of the phase diagram and microstructure. In addition, it predicts a variant of this phase that does not seem to have been experimentally observed. Using this model we have calculated generic phase diagrams of two-component membranes. We have also determined the phase diagram of a two-component lipid membrane from x-ray diffraction studies on aligned multilayers. This phase diagram is found to be in good agreement with that calculated from the model.

DOI: 10.1103/PhysRevE.85.051701

PACS number(s): 61.30.Dk, 87.16.D–

**I. INTRODUCTION**

Phospholipids are amphiphilic molecules with a hydrophilic head group and one or more hydrophobic hydrocarbon chains, and are the major constituents of biomembranes. They self-assemble in water to form a variety of thermodynamic phases, depending upon the temperature and water content [1]. The most common of which is the lamellar phase, a periodic structure consisting of lipid bilayers separated by layers of water.

Above the chain-melting transition (also known as the main transition) temperature  $T_m$ , phospholipids form the fluid  $L_\alpha$  phase, where the hydrocarbon chains are disordered with many trans-gauche isomerizations in their C-C bonds. Positional order in the plane of the bilayer is liquidlike in this phase. The lower temperature gel ( $L_\beta$  or  $L_{\beta'}$ ) phase is characterized by flat bilayers with the hydrocarbon chains predominantly in the fully stretched all-trans conformation. In the  $L_{\beta'}$  phase the chains are tilted relative to the bilayer normal, whereas in the  $L_\beta$  phase there is no such tilt. The presence of a tilt depends on the relative cross-sectional areas of the head group and chains of the lipid species [2,3]. For example, phosphatidylcholines display a tilt angle of about  $30^\circ$  while phosphatidylethanolamines with relatively smaller head groups do not show any tilt.

Phospholipids such as phosphatidylcholines (PCs) exhibit a modulated phase between the  $L_\alpha$  and the  $L_{\beta'}$  phases at high water content [1,4–6]. This is known as the  $P_{\beta'}$  or the ripple phase, and is characterized by a one-dimensional (1D) periodic height modulation of the bilayers. In this phase the hydrocarbon chains are partially ordered and have a nonzero tilt relative to the bilayer normal. The first-order  $L_{\beta'}$ - $L_\alpha$  transition is well understood in terms of the melting of some of the degrees of freedom within the hydrocarbon chains [7]. However, the first-order  $L_{\beta'}$ - $P_{\beta'}$  transition (often called the pretransition), which occurs at a temperature slightly below the main transition, is much less understood.

The structural features of the ripple phase have been extensively studied using a variety of techniques such as x-ray diffraction (XRD) [1,8–11], neutron diffraction [12], freeze-fracture electron microscopy [8,13–16], and scanning-tunneling microscopy [9,17]. These experiments have revealed the existence of both symmetric as well as asymmetric ripples. The asymmetric structure is, however, thermodynamically more stable and is commonly reported in almost all x-ray studies on PC systems. The wavelength of the ripples typically lies in the range of 12–16 nm [18] and the peak-to-peak amplitude is about 2 nm. X-ray diffraction patterns from this phase can be indexed on a two-dimensional oblique lattice and the rippled bilayers lack a mirror plane normal to the rippling direction. Electron density maps calculated from the diffraction data [11,19–22] confirm that the bilayers have a sawtoothlike height profile, with the bilayer thickness appreciably different in the two arms (i.e., the two segments that make up each tooth) of the ripple. The absence of a mirror plane normal to the ripple wave vector in the asymmetric ripple phase can be attributed to the presence of asymmetry in either the shape (i.e., one arm being longer than the other) or bilayer thickness (with a higher bilayer thickness in one of the arms) or both. However, in almost all the systems studied so far both these features are present simultaneously.

Several theoretical models have been proposed to describe the ripple phase [23–30], but none of them account for all the experimental observations, namely, (i) occurrence of the  $P_{\beta'}$  phase between  $L_\alpha$  and  $L_{\beta'}$  phases, separated by two first-order transitions; (ii) unequal bilayer thickness in the two arms of the ripple; and (iii) unequal lengths of the two arms.

Macroscopic continuum theories, which treat the bilayer as a continuous membrane, can broadly be classified into two main categories depending on the nature of the order parameter that is chosen to describe the ripple phase. One set of theories considers the order parameter to be a scalar, such as the membrane thickness as in the models by Falkovitz *et al.* [24], Marder *et al.* [25], and by Goldstein and Leibler (GL) [27,28],

or the configuration of the hydrocarbon chains (ratio of cis to trans bonds in the chains) [31]. Due to the scalar nature of the order parameter, these models cannot distinguish the different symmetries of the  $L_\alpha$ ,  $L_{\beta'}$ , and  $P_{\beta'}$  phases. Further, in these models the ripple phase is characterized by a bilayer thickness modulation and not a height modulation. The other set of continuum theories considers the order parameter to be a vector, such as the tilt of the hydrocarbon chains with respect to the local layer normal, as in the Lubensky-MacKintosh (LM) [29] and the Seifert-Shillcock-Nelson models [32].

Although models involving vectorial order parameter can predict the various phases with different in-plane symmetries, they have their own limitations. For example, the LM model cannot account for the occurrence of the ripple phase in bilayers composed of achiral molecules and speculated that the the origins of the experimentally observed asymmetric ripples lay in the chirality of lipid molecules. However, subsequent experiments using racemic (achiral) mixtures showed this was not the case [33,34].

In this paper we present a phenomenological Landau model to describe the ripple phase, that unifies the two distinct approaches (those using scalar and vector order parameters) used in the earlier models. In addition to the coupling between the curvature of the bilayer to the divergence of the molecular tilt [29], we bring out the importance of the coupling of variation in the bilayer thickness to molecular tilt and show that: (i) the asymmetry of the ripple phase arises from (a) the difference in the bilayer thickness in the two arms of the ripple, and (b) the unequal length of the two arms; and (ii) the existence of a nonzero mean tilt of the hydrocarbon chains of the lipid molecules along the direction of ripple.

In Sec. II we present the model for single-component achiral membranes. The main results of this section have already been reported [35]. In Sec. III we extend the model to two-component membranes and use it to calculate the phase diagrams for various binary lipid mixtures. In Sec. IV we present the phase diagram of dipalmitoyl phosphatidylcholine (DPPC) - dimyristoyl phosphatidylcholine (DMPC) mixtures determined from small-angle x-ray scattering studies of aligned samples. The experimental phase diagram is found to be in good agreement with the one calculated using our model.

## II. SINGLE-COMPONENT ACHIRAL MEMBRANES

In this section, we develop a phenomenological model for describing the asymmetric ripple phase in achiral membranes. To simplify matters, we ignore interbilayer interactions and choose to describe the ripple phase in terms of (i) a scalar order parameter  $\psi$ , which represents the relative bilayer thickness, and (ii) a vector order parameter  $\mathbf{m}$ , which is the projection of the molecular axis in the plane tangent to the bilayer surface (Fig. 1) [29]. We consider three distinct contributions to the total free energy density:

### A. The stretch-free energy density

The main transition is characterized by the freezing of the lateral motion as well as conformational ordering of the hydrocarbon chains. Many experiments have revealed

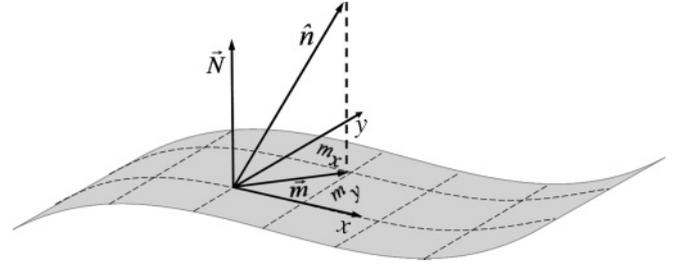


FIG. 1. The unit vector  $\hat{\mathbf{n}}$  represents the orientation of the long axis of the lipid molecules relative to  $\mathbf{N}$ , the bilayer normal. The projection of  $\hat{\mathbf{n}}$  on the bilayer plane  $\mathbf{m} [= \hat{\mathbf{n}} - (\mathbf{N} \cdot \hat{\mathbf{n}}) \mathbf{N}]$  is the order parameter.  $m_x$  and  $m_y$  are the components of  $\mathbf{m}$  along the two orthogonal directions in the plane of the bilayer.

that at the main transition there is a discontinuous jump in the membrane thickness [36]. Therefore, following GL, we describe the main transition as one involving a change in the bilayer thickness and introduce a dimensionless scalar order parameter  $\psi$ , such that

$$\psi = \frac{\delta(\mathbf{r}) - \delta_0}{\delta_0}, \quad (1)$$

where  $\delta(\mathbf{r})$  is the actual membrane thickness,  $\delta_0$  is the constant thickness of the membrane in the  $L_\alpha$  phase and  $\mathbf{r}$  denotes the lateral position within the bilayer plane. No specific assumption is made about the microscopic state of the lipids in choosing  $\psi$  as the order parameter; we can think of  $\psi$  as a hybrid coarse-grained variable that summarizes changes in various types of degrees of freedom, such as conformational changes of the hydrocarbon chains.  $\psi$  is taken to be positive for  $T < T_m$  due to the stretching of the chains. This is valid in general, even if the chains are tilted below  $T_m$ .

The stretching free energy per unit area, for an isolated lipid bilayer membrane, can therefore be written as [37]

$$f_{st} = \frac{1}{2} a_2 \psi^2 + \frac{1}{3} a_3 \psi^3 + \frac{1}{4} a_4 \psi^4 + \frac{1}{2} C (\nabla \psi)^2 + \frac{1}{2} D (\nabla^2 \psi)^2 + \frac{1}{4} E (\nabla \psi)^4. \quad (2)$$

The explicit temperature dependence is assumed to reside solely in the coefficient of  $\psi^2$

$$a_2 = a'_2 (T - T^*),$$

$T^*$  being a reference temperature (which in the absence of the cubic term in  $f_{st}$  is the critical temperature). Since,  $L_{\beta'} \rightarrow L_\alpha$  phase transition is known to be first order, the coefficient  $a_3$  is taken to be negative. In the presence of the cubic term, the critical temperature  $T^*$  is preempted by a first-order melting transition at,

$$T_m = T^* + \frac{2a_3^2}{9a'_2 a_4}. \quad (3)$$

The coefficient  $C$  can either be positive or negative but  $D$ ,  $E$ , and  $a_4$  are always positive to ensure the stability of this free energy expansion. With  $C > 0$ , the equilibrium phases are always spatially homogeneous; either as  $L_\alpha$  or  $L_\beta$  ( $L_{\beta'}$ ). However, with  $C < 0$  modulated phases are possible with some characteristic wave vector  $q_0$ . To motivate how  $C$  can be negative, it is convenient to introduce an auxiliary scalar variable  $\rho$ . Constructing effective cylinders enclosing the head

group and the chain with cross-sectional areas  $a_h$  and  $a_t$ , respectively, we define  $\rho \equiv (a_h - a_t)/(a_h + a_t)$ . Consider a bilayer membrane whose thickness  $\psi$  increases steadily over a length scale  $\xi$ . This variation in thickness produces a strain at the molecular level. This can be accommodated both by a variation in the tilt  $\mathbf{m}$ , and a variation in the mismatch  $\rho$  over this scale. The latter coupling to lowest order is of the form  $\nabla\psi \cdot \nabla\rho$ . This term, consistent with the symmetries of the bilayer, will generically lead to  $C < 0$ .

### B. The tilt free energy density $f_{\text{tilt}}$

The tilt free energy density can then be written in terms of  $\mathbf{m}$  as,

$$\begin{aligned} f_{\text{tilt}} = & \frac{1}{2}b_2|\mathbf{m}|^2 + \frac{1}{4}b_4|\mathbf{m}|^4 + \Gamma_1(\nabla \cdot \mathbf{m})^2 \\ & + \Gamma_2(\nabla^2\mathbf{m})^2 + \Gamma_3(\nabla \cdot \mathbf{m})^4 + \Gamma_4\psi|\mathbf{m}|^2 \\ & + \Gamma_5(\mathbf{m} \cdot \nabla\psi)^2 + \Gamma_6(\mathbf{m} \times \nabla\psi)^2 \\ & + \Gamma_7(\nabla^2\psi)(\nabla \cdot \mathbf{m})^2 + \Gamma_8(\nabla\psi)^2(\nabla \cdot \mathbf{m})^2. \end{aligned} \quad (4)$$

First and second terms in the above equation represent the coupling of the molecular tilt vector  $\mathbf{m}$  to the bilayer thickness and modulations in the thickness, respectively. The third term corresponds to the splay deformation of the  $\mathbf{m}$  field and represent the tilt elasticity in the tilted phase. Since the order parameter  $\mathbf{m}$  is a vector, cubic terms in  $\mathbf{m}$  are not allowed.

As shown by Jacobs *et al.* [38] in the context of a one-dimensional model with a scalar order parameter, a term in the fourth power of the gradient of the order parameter is necessary to stabilize a modulated phase with a nonzero mean value of the order parameter. Hence we include  $(\nabla\psi)^4$  and  $(\nabla \cdot \mathbf{m})^4$  terms in our expression for stretching free energy density and the tilt free energy density, respectively.

As  $b_2$  is positive in our model, the term involving the coefficient  $\Gamma_4$ , which couples the molecular tilt to the change in the bilayer thickness, is the most crucial one for the appearance of the asymmetric ripple phase. When  $\Gamma_4 > 0$ , the stable phase below  $T_m$  is  $L_\beta$  with  $|\mathbf{m}| = 0$ . On the other hand, tilted phases can form if  $\Gamma_4 < 0$ . The succeeding two terms take into account the anisotropy of the tilted bilayer. The next two terms represent higher order couplings between modulations in  $\psi$  and in  $\mathbf{m}$ , allowed by the symmetry of the system. These higher order coupling terms are kept merely for consistency and are not crucial for obtaining the optimal phases.

### C. The curvature energy density $f_c$

The curvature energy density of the bilayer can be written as [29]

$$f_c = \frac{1}{2}\kappa(\nabla^2 h)^2 - \gamma(\nabla^2 h)(\nabla \cdot \mathbf{m}), \quad (5)$$

where  $h(x,y)$  is the height of the bilayer relative to a flat reference plane,  $\kappa$  is the bending rigidity of the membrane, and the coefficient  $\gamma$  couples the mean curvature to splay in  $\mathbf{m}$ .

The total free energy density is given by the sum of the above three contributions

$$f = f_{\text{st}} + f_{\text{tilt}} + f_c. \quad (6)$$

The equilibrium height profile of the bilayer  $h(x,y)$  is related to the tilt  $\mathbf{m}$  via the Euler-Lagrange equation,

$$\nabla^2 h = \frac{\gamma}{\kappa}(\nabla \cdot \mathbf{m}). \quad (7)$$

Eliminating  $h$  from the free energy density  $f$  leads to the effective energy density  $f_{\text{eff}}$  with a reduced splay elastic constant.

$$\tilde{\Gamma}_1 = \Gamma_1 - \frac{\gamma^2}{\kappa}.$$

Thus, the effective free energy density is given by,

$$\begin{aligned} f_{\text{eff}} = & \frac{1}{2}a_2\psi^2 + \frac{1}{3}a_3\psi^3 + \frac{1}{4}a_4\psi^4 + \frac{1}{2}C(\nabla\psi)^2 \\ & + \frac{1}{2}D(\nabla^2\psi)^2 + \frac{1}{4}E(\nabla\psi)^4 + \frac{1}{2}b_2|\mathbf{m}|^2 \\ & + \frac{1}{4}b_4|\mathbf{m}|^4 + \tilde{\Gamma}_1(\nabla \cdot \mathbf{m})^2 + \Gamma_2(\nabla^2\mathbf{m})^2 \\ & + \Gamma_3(\nabla \cdot \mathbf{m})^4 + \Gamma_4\psi|\mathbf{m}|^2 + \Gamma_5(\mathbf{m} \cdot \nabla\psi)^2 \\ & + \Gamma_6(\mathbf{m} \times \nabla\psi)^2 + \Gamma_7(\nabla^2\psi)(\nabla \cdot \mathbf{m})^2 \\ & + \Gamma_8(\nabla\psi)^2(\nabla \cdot \mathbf{m})^2. \end{aligned} \quad (8)$$

To determine the mean-field phase diagram we assume that the modulated phase is characterized by the most dominant wave vector  $\mathbf{q}$ , and choose the following ansatz for  $\psi$  and  $\mathbf{m}$ ,

$$\begin{aligned} \psi &= \psi_0 + \psi_1 \sin(qx) \\ m_x &= m_{0x} + m_{1x} \cos(qx) + m_{2x} \sin(qx) \\ &+ m_{3x} \cos(2qx) + m_{4x} \sin(2qx) \\ m_y &= m_{0y} \end{aligned} \quad (9)$$

with the amplitudes  $\{\psi_i, m_i\}$  as variational parameters. We retain Fourier components of  $m_x$  to second order to account for the ripple asymmetry; the experimentally observed asymmetric ripple profile is obtained if  $m_{3x} \neq 0$ . In principle, one could have done a numerical variational calculation with many more Fourier components. However, our simplified ansatz is sufficient to recover all the qualitative features of the experimentally observed asymmetric ripple phase.

2D modulations of the membrane [39] were not included in our ansatz [Eq. (9)], since earlier studies [21,29,30] based on a similar free energy density for the tilt and curvature alone, show that the presence of a mean tilt suppresses two-dimensional height modulations. The additional  $\psi$ -dependent terms in the free energy density do not alter this conclusion. Hence we confine our attention to one-dimensionally modulated ripples. Note that spatial modulations in  $m_y$  are neglected as we do not keep terms proportional to  $(\nabla \times \mathbf{m})$  in Eq. (4). Variation of  $m_y$  with  $x$  represents nonparallel orientations of the lipid chains, which are not favored by van der Waal's interactions between the chains. Such terms are important in chiral systems, where they can lead to structures with nonzero winding numbers [29]. In the absence of chirality, these terms can, however, be neglected.

The average free energy is obtained by integrating the  $F_{\text{eff}}$  over one spatial period, i.e.,

$$\langle f_{\text{eff}} \rangle = \frac{\int_0^{\frac{2\pi}{q}} f_{\text{eff}} dx}{\int_0^{\frac{2\pi}{q}} dx}. \quad (10)$$

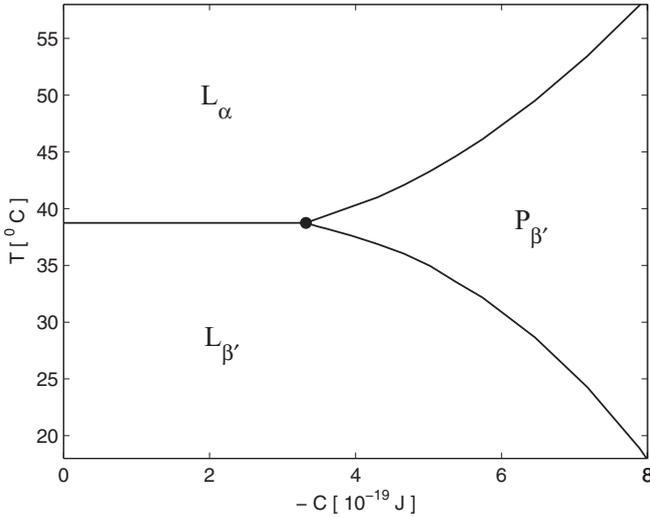


FIG. 2. Phase diagram in the  $T$ - $C$  plane calculated from the model.  $a'_2 = 159.42 k_B$ ,  $T^* = 260.0$  K. Values of the other coefficients in units of  $k_B T^*$  are:  $a_3 = -306.5$ ,  $a_4 = 613.15$ ,  $b_2 = 0.2$ ,  $b_4 = 200.0$ ,  $D = 557.41$ ,  $E = 600.0$ ,  $\Gamma_1 = 0.010$ ,  $\Gamma_2 = 1.80$ ,  $\Gamma_3 = 500.0$ ,  $\Gamma_4 = -3.0$ ,  $\Gamma_5 = -20.0$ ,  $\Gamma_6 = -20.0$ ,  $\Gamma_7 = -50.0$ ,  $\Gamma_8 = -25.0$ . Both the main transition ( $L_\alpha \rightarrow L_{\beta'}$ ;  $L_\alpha \rightarrow P_{\beta'}$ ) and pretransition ( $P_{\beta'} \rightarrow L_{\beta'}$ ) are first order.

#### D. Results and discussion

The phase diagram in the  $C$ - $T$  plane is obtained from the numerical minimization of the effective free energy  $\langle f_{\text{eff}} \rangle$ . Figure 2 represents an example of such a phase diagram. It is calculated for a choice of parameter values, which reproduce closely the main and pretransition temperatures of DPPC at  $C = -4.84 \times 10^{-19} J$ . These values (except for  $\Gamma_1$  to  $\Gamma_6$ ) are taken from Ref. [28] and Ref. [37]. It is seen from Fig. 2 that there are three distinct regions in the phase diagram corresponding to three different phases

$$\begin{aligned}
 L_\alpha \text{ phase : } & \begin{cases} \psi_0 = 0, & \psi_1 = 0, & m_{0x} = 0, & m_{0y} = 0 \\ m_{1x} = 0, & m_{2x} = 0, & m_{3x} = 0, & m_{4x} = 0 \end{cases} \\
 L_{\beta'} \text{ phase : } & \begin{cases} \psi_0 \neq 0, & \psi_1 = 0, & m_{0x} \neq 0, & m_{0y} \neq 0 \\ m_{1x} = 0, & m_{2x} = 0, & m_{3x} = 0, & m_{4x} = 0 \end{cases} \\
 P_{\beta'} \text{ phase : } & \begin{cases} \psi_0 \neq 0, & \psi_1 \neq 0, & m_{0x} \neq 0, & m_{0y} = 0 \\ m_{1x} = 0, & m_{2x} \neq 0, & m_{3x} \neq 0, & m_{4x} = 0. \end{cases}
 \end{aligned}$$

The first-order transition lines which separate these three phases meet at a Lifshitz point at  $C_{\text{Lp}} = -3.43 \times 10^{-19} J$  and  $T = 38.75$  °C. When  $|C| < |C_{\text{Lp}}|$ , the first-order transition line is parallel to the  $C$  axis and occurs at  $T = 38.75$  °C. As long as  $|C| < |C_{\text{Lp}}|$  the  $L_{\beta'}$  phase melts directly to the  $L_\alpha$  phase at  $T_m$ . But for  $|C| > |C_{\text{Lp}}|$  the  $L_{\beta'}$  phase first melts into the  $P_{\beta'}$  phase, which in turn transforms into the  $L_\alpha$  phase at higher temperatures. It is also clear from the figure that, as  $|C|$  increases, the region of the  $P_{\beta'}$  phase expands at the expense of the  $L_{\beta'}$  and the  $L_\alpha$  phases.

It would seem that our free energy functional contains a large number of parameters, which we need to fine tune to obtain the asymmetric ripple phase. We would like to emphasize that asymmetric ripples are a fairly generic and robust feature of our model whenever  $\Gamma_4 < 0$ . The other  $\Gamma_i$

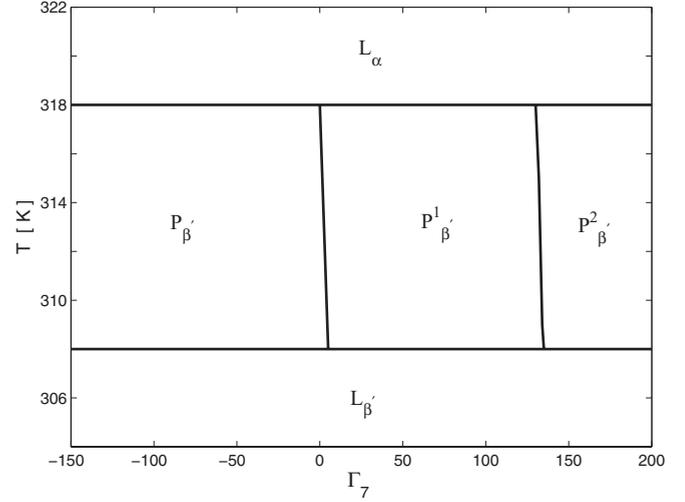


FIG. 3. Dependence of the phase behavior on  $\Gamma_7$ . All lines in the phase diagram correspond to first-order transitions.  $C = 5.74 \times 10^{-19} J$ , and all other parameters as in Fig. 2.

only make the shape anisotropy in the height profile more pronounced when they are negative, and are not crucial for the existence of the asymmetric ripple phase. Further, x-ray diffraction experiments identify the asymmetric ripple phase on the basis of its oblique unit cell. Such a unit cell can arise either from a strongly asymmetric height profile or from a weakly asymmetric height profile as long as the thickness is different in the two arms of the ripple [21]. This feature is exactly what is captured within our theory [Figs. 4(a) and 4(b)].

In addition to  $C$  and  $\Gamma_4$ , the phase behavior depends crucially on  $\Gamma_7$ . Depending on the value of  $\Gamma_7$  three distinct modulated phases are found, with differing spatial variations of the order parameters (Fig. 3). For  $\Gamma_7 < 0$ , the  $P_{\beta'}$  phase is stable, where the height profile is asymmetric and resembles very closely those seen in experiments [Fig. 4(a)] [17,18,40]. In this phase  $m_{1x} = 0$ ,  $m_{2x} \neq 0$ ,  $m_{3x} \neq 0$ ,  $m_{4x} = 0$ .  $\psi_1$  is almost  $\pi/2$  out of phase with  $h$ , so that it is positive (negative) along the longer (shorter) arm of the ripple, resulting in different bilayer thicknesses in the two arms, again in agreement with experimental observations [18,40]. When  $\Gamma_7$  is weakly positive, the  $P_{\beta'}^1$  is obtained [Fig. 4(b)], where the higher Fourier components of  $m_x$  are absent ( $m_{1x} = 0$ ,  $m_{2x} \neq 0$ ,  $m_{3x} = 0$ ,  $m_{4x} = 0$ ) and hence the height profile is symmetric. As in the  $P_{\beta'}$  phase the bilayer thickness is different in the two arms. This profile resembles the structure reported in Ref. [21], based on the diffraction data of Ref. [1]. For higher values of  $\Gamma_7$  the  $P_{\beta'}^2$  is found [Fig. 4(c)], where  $m_{1x} \neq 0$ ,  $m_{2x} = 0$ ,  $m_{3x} = 0$ ,  $m_{4x} \neq 0$ . The height profile is symmetric and is almost in phase with  $\psi_1$ . As a result, the bilayer thickness is modulated within each arm of the ripple. This structure does not seem to have been observed in any experiments until now. However, since the electron density map of the ripple phase has been calculated only in a few instances, the possible occurrence of this structure in some systems cannot be ruled out.

The present model predicts the chain tilt to be high in the thicker arm and negligible in the thinner one. There are in

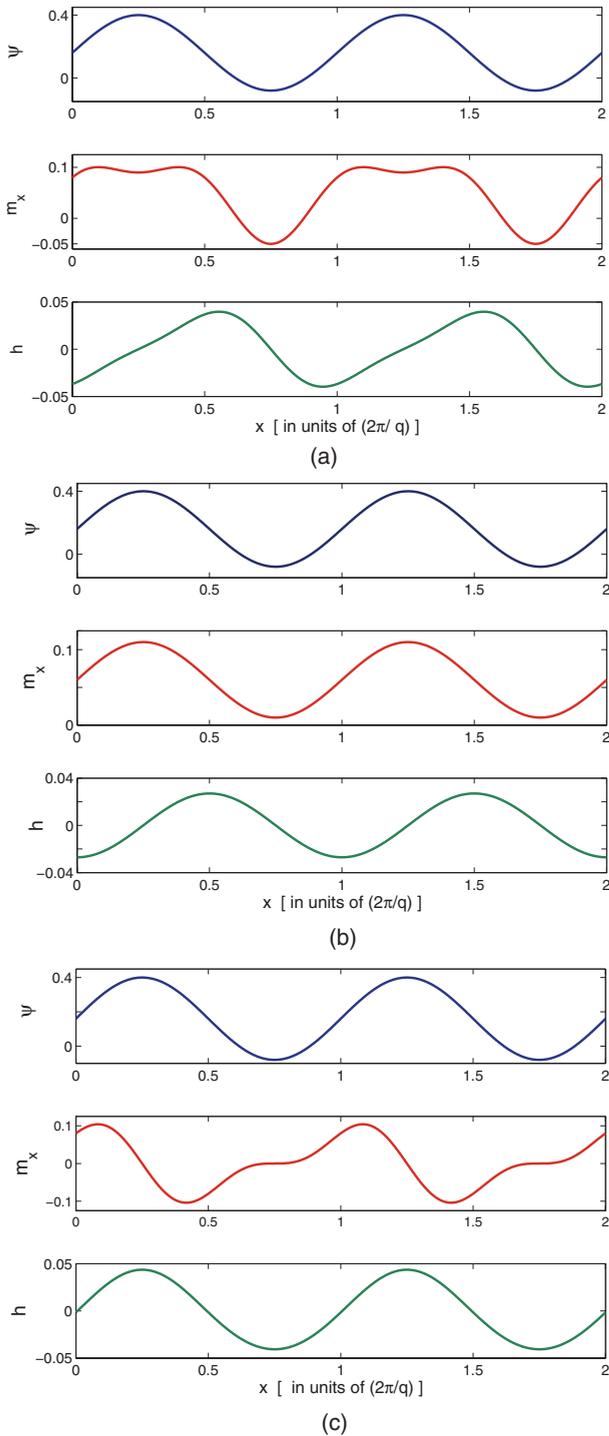


FIG. 4. (Color online) Spatial variation of  $\psi$ ,  $m_x$ , and  $h$  in the different ripple phases obtained for (a)  $\Gamma_7 = -150 k_B T^*$ , (b)  $\Gamma_7 = 100 k_B T^*$ , and (c)  $\Gamma_7 = +150 k_B T^*$ .  $T = 310.50$  K and all other parameters are the same as those used in Fig. 2.

principle at least two chain conformations consistent with this thinner arm of the bilayer: one is by having disordered chains, the other by chain interdigitation. Since in our model, the high-temperature phase was identified as  $L_\alpha$ , it is natural to populate the thinner arm with disordered chains. We therefore propose that in the asymmetric ripple, the thicker arm is made up of tilted gel-like domains and the thinner one is

fluidlike with negligible chain tilt. This conclusion is partially supported by spectroscopy and diffusion experiments, which indicate a significant fraction of disordered chains in the ripple phase [41,42]. However, it must be acknowledged that there is no direct experimental data on the spatial distribution of the disordered chains within the rippled bilayer. Electron density maps calculated from diffraction data do not provide any direct information about chain ordering. In contrast, recent computer simulation studies of the ripple phase using a variety of model lipids at different levels of coarse-graining [43–45], find an asymmetric ripple structure consistent with the height and thickness modulations seen in the electron density maps, with the thicker arm consisting of gel-like domains, and the thinner one made up of ordered and interdigitated chains. The crossover region between these two contains a large fraction of disordered chains. In the gel-like domains chains are tilted with respect to the local layer normal, whereas chain tilt vanishes in the thinner arm due to interdigitation. Thus the height, thickness, and tilt modulations found in these simulations are consistent with the predictions of the present model, although the thinner arm consists of interdigitated chains instead disordered chains as proposed here. Symmetric ripples consisting of alternating gel-like and fluidlike domains have also been found in some simulations [46]. These ripples are characterized by only a thickness modulation and no height modulation. Stacks of such bilayers will result in a centered rectangular lattice and do not seem to have been observed in any pure lipid-water system.

### III. BINARY LIPID MIXTURES

Biomembranes are multicomponent, including lipid species that show liquid-ordered behavior, and are likely to have thickness variations [47]. The first step in understanding such complex systems is to study binary mixtures of lipids, which differ in their head groups and chains. Therefore, in this section, we extend the model presented in the previous section for single-component membranes to membranes consisting of two lipid species.

We specifically consider two different cases. In case 1, only one of the lipids exhibits the ripple phase. Since the occurrence of the ripple phase is closely related to the formation of a tilted gel phase at lower temperatures, one of the lipids is assumed to show the phase sequence  $L_\alpha \rightarrow P_{\beta'} \rightarrow L_{\beta'}$ , and the other to show  $L_\alpha \rightarrow L_\beta$  on cooling. This case corresponds to two species differing only in their head groups. In case 2, both the lipids exhibit the ripple phase, and corresponds to the situation where the two lipids differ only in the length of their hydrocarbon chains. We employ an approach quite similar to that used in Ref. [37] and ignore any lipid exchange with the surrounding solvent. Owing to the fact that the lipids constituting the mixture can have different chain lengths, degrees of saturation, or head groups, the main transition temperature is taken to be different for the two lipids.

Due to the presence of an additional component in the system there is a contribution  $f_{\text{mix}}$  to the total free energy, corresponding to the free energy of mixing.  $f_{\text{mix}}$  is given by the sum of the entropy of mixing and enthalpy, which within the Bragg-Williams (mean-field) approximation can be written

as,

$$f_{\text{mix}} = k_B T [\phi \log(\phi) + (1 - \phi) \log(1 - \phi)] + \frac{1}{2} J \phi (1 - \phi) \quad (11)$$

where,  $k_B$  is the Boltzmann constant, and  $J > 0$  is an attractive interaction parameter that enhances lipid-lipid demixing.

Since the important properties of the lipids are mainly reflected in the model parameters  $T^*$ ,  $C$ , and  $\Gamma_4$ , we assume their values for a mixture to be the average of that for the two pure lipids weighted by their concentration

$$C(\phi) = \phi C_A + (1 - \phi) C_B \quad (12)$$

$$\Gamma_4(\phi) = \phi \Gamma_{4A} + (1 - \phi) \Gamma_{4B} \quad (13)$$

$$T^*(\phi) = \phi T_A^* + (1 - \phi) T_B^* \quad (14)$$

where, the subscripts  $A$  and  $B$  refer to the lipids  $A$  and  $B$ , respectively, and  $\phi$  is the mole fraction of lipid  $A$  in the mixture. All other parameters are assumed to be same for both the lipids.

Combining Eqs. (8) and (11), we obtain the total free energy density  $f_{\text{total}}$  as,

$$f_{\text{total}} = f_{\text{mix}} + f_{\text{eff}}. \quad (15)$$

After minimizing Eq. (15) with respect to both  $\psi$  and  $\mathbf{m}$ , the two-phase coexistence in the  $(T, \phi)$  plane is obtained using Maxwell's construction.

## A. Results and discussion

### 1. Tilt-nontilt mixtures

We now calculate the phase diagram of binary lipid mixtures, where only one of the constituents exhibits the rippled phase, such as mixtures of dipalmitoyl phosphatidylcholine (DPPC) and dipalmitoyl phosphatidylethanolamine (DPPE). The phosphatidylcholines, which have the  $-N(\text{CH}_3)_3^+$  moiety in their polar head groups, typically exhibit a tilted  $L_{\beta'}$  phase. On the other hand, the phosphatidylethanolamines, which have the smaller  $-(\text{NH}_3)_3^+$  moiety instead, shows the untilted  $L_{\beta}$  phase and do not exhibit the ripple phase.

From Fig. 2 one finds that only when the value of  $|C|$  is greater than  $|C_{Lp}|$ , a spatially modulated phase occurs between the high- and low-temperature phases. The sign of  $\Gamma_4$  is also important in determining the existence of a tilted phase at lower temperatures. If  $\Gamma_4 < 0$ , then the tilted phase  $L_{\beta'}$  occurs, otherwise one has the untilted  $L_{\beta}$  phase below the main transition temperature.

Keeping this in mind, we choose the value of  $|C|$  for lipid  $A$  (DPPE) to be less than  $|C_{Lp}|$  and that for lipid  $B$  (DPPC) to be greater than  $|C_{Lp}|$ . The parameter values are chosen so as to reproduce the transition temperatures of DPPC and DPPE.  $\Gamma_4$  is chosen to be negative for DPPC and positive for DPPE.

Using the above values of the parameters the binary phase diagram for DPPC-DPPE mixtures were calculated, and the results are shown in Figs. 5 and 6. In Fig. 5 the interaction parameter  $J = 4.0 k_B T^*$  while in Fig. 6  $J = 5.0 k_B T^*$ .

The calculated phase diagram of Fig. 5 is found to be in good quantitative agreement with the experimental one for DPPC-DPPE mixtures [48], and is very similar to what is obtained for other lipid mixtures such as, DMPC- dipalmitoyl

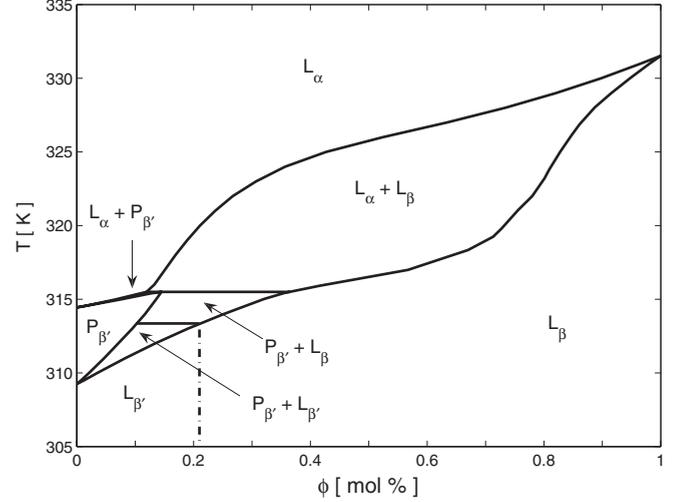


FIG. 5. Calculated mean-field phase diagram of DPPC-DPPE mixture as a function of composition and temperature  $T$ .  $\phi$  is the mole fraction of DPPE.  $C_{\text{DPPC}} = 4.66 \times 10^{-19} J$ ,  $C_{\text{DPPE}} = 3.58 \times 10^{-21} J$ ,  $T_{\text{DPPC}}^* = -17^\circ \text{C}$ ,  $T_{\text{DPPE}}^* = -3^\circ \text{C}$ , and  $J = 4.0 k_B T^*$ . All other parameter values are as in Fig. 2.

phosphatidylserine (DPPS), and DPPC-DPPS [8]. At higher temperatures, we have the  $L_{\alpha}$  phase for the entire range of composition. With decreasing temperature the  $P_{\beta'}$  phase appears very close to the pure DPPC axis, and is bounded by coexistence regions with the  $L_{\beta'}$ ,  $L_{\beta}$ , and the  $L_{\alpha}$  phases. The  $P_{\beta'}$  -  $L_{\alpha}$  coexistence region is very narrow on the scale of the figure and collapses almost into a line. At low temperatures there is a continuous  $L_{\beta'}$  -  $L_{\beta}$  transition at  $\phi \sim 0.2$ , which reflects the change in the sign of  $\Gamma_4$  according to Eq. (14).

Figure 6 shows the phase diagram for a binary lipid system in which the constituent lipids show a very high tendency to segregate and resembles the experimentally obtained one for dielaidoyl phosphatidylcholine (DEPC)-DPPE mixtures [49]. The two lipids are completely miscible only in the

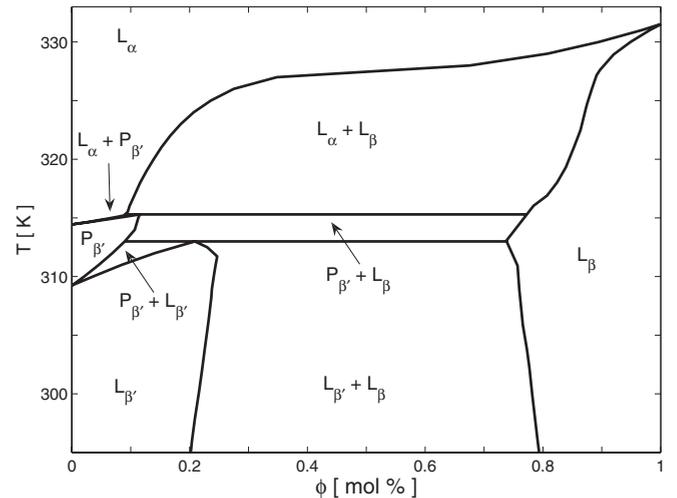


FIG. 6. Calculated mean-field phase diagram of a binary lipid mixture as a function of composition and temperature at  $J = 5.0 k_B T^*$ . All other parameter values are as in Fig. 5.

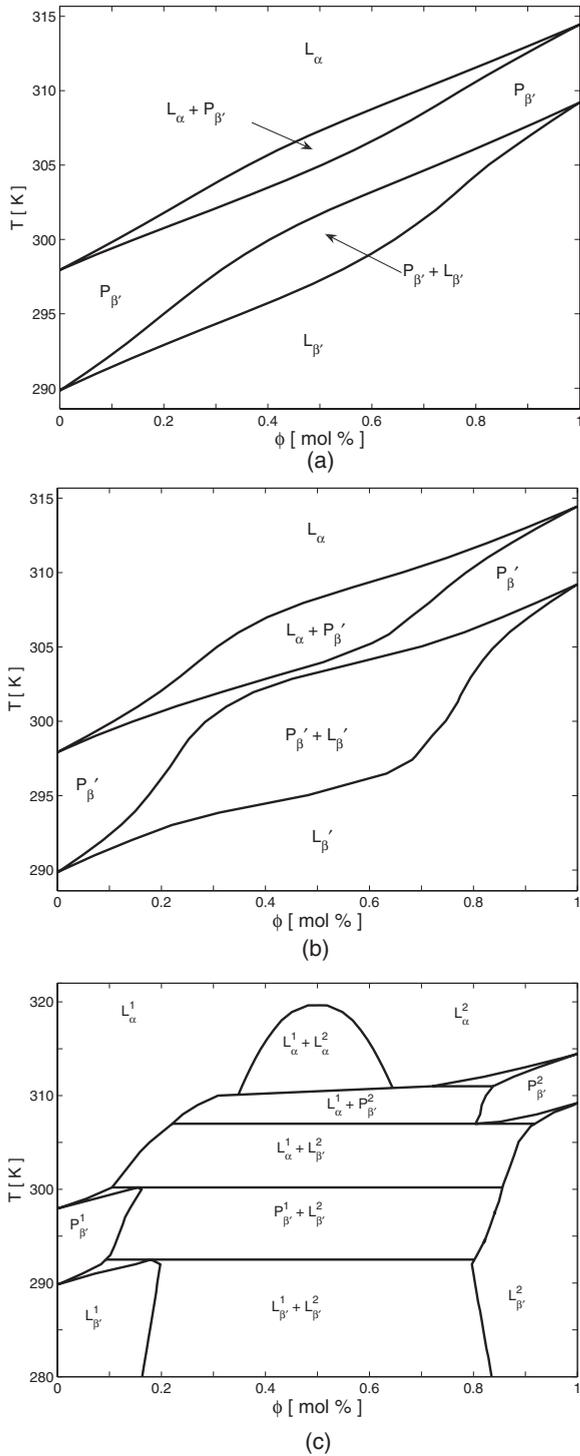


FIG. 7. Calculated mean-field phase diagram of a binary lipid mixture as a function of their relative composition  $\phi$  and temperature  $T$ . Both the lipids constituting the mixture exhibit a low-temperature tilted phase. The lipid interaction parameter is set to be (a)  $J = 3.0 k_B T^*$ , (b)  $J = 4.0 k_B T^*$ , (c)  $J = 5.0 k_B T^*$ .  $C_{DMPC} = 5.20 \times 10^{-19} J$  and  $C_{DPPC} = 4.66 \times 10^{-19} J$ . All other parameter values are as in Fig. 2.

$L_\alpha$  phase. The main qualitative difference from the previous case is the large region of  $L_{\beta'} - L_\beta$  coexistence at lower temperatures.

### 2. Tilt-tilt mixtures

In this section, we consider the case where both the constituents exhibit the ripple phase between the  $L_\alpha$  and  $L_{\beta'}$  phases. This would correspond to mixtures of lipids such as DPPC and DMPC. We, therefore, choose the value of  $C$  for the two lipids to be larger than  $C_{LP}$  and the sign of  $\Gamma_4$  to be negative for both of them. The reference temperature for A and B is chosen to be,  $T_A^* = -35^\circ\text{C}$  and  $T_B^* = -17^\circ\text{C}$ , respectively. The value of  $\Gamma_4$  is taken to be the same for both the lipids. Using these values of the parameters the binary phase diagram was calculated, and the results are shown in Figs. 7(a)–7(c) for different values of  $J$ .

When  $J = 3.0 k_B T^*$  [Fig. 7(a)] the three phases extend throughout the entire range of composition. These regions are separated by two coexistence regions of  $L_\alpha - P_{\beta'}$  and  $P_{\beta'} - L_{\beta'}$  phases. As  $J$  is increased to  $4.0 k_B T^*$  [Fig. 7(b)], these coexistence regions almost touch each other, confining the  $P_{\beta'}$  phase to a narrow strip. With further increase in the value of  $J(=5.0 k_B T^*)$  two-phase coexistence regions dominate the phase diagram with the pure phases confined to compositions close to the two pure lipids. For this value of  $J$  the two lipids show a miscibility gap in the  $L_\alpha$  phase, with the appearance of a critical point. As discussed in the next section, the calculated phase behavior at low values of  $J$  closely resembles that observed in mixtures of very similar lipids. Although the complex phase diagrams calculated for high values of  $J$  have not yet been reported in experiments, we believe that such phase behavior should be seen in binary mixtures of very dissimilar lipids, such as those with hydrocarbon and fluorocarbon chains.

### IV. PHASE BEHAVIOR OF DPPC-DMPC MIXTURES

Small-angle scattering techniques can in principle detect microscopic phase separation in the plane of bilayers, if there is sufficient contrast in the scattering densities of the two phases. However, even in the absence of such contrast, macroscopic phase separation can easily be detected from non overlapping reflections in the diffraction pattern coming from the individual phases. On the basis of the diffraction patterns we have determined the partial phase diagram of the binary mixture consisting of DPPC and DMPC. The use of oriented samples has an advantage that the in-plane ordering of molecules can be easily inferred from the wide-angle reflections. For example, in the gel phase of both DPPC and DMPC, one observes two wide-angle reflections, one on-axis ( $q_z = 0$ ) and the other off-axis ( $q_z \neq 0$ ), coming from the quasihexagonal lattice of hydrocarbon chains of the lipid molecules. The positions of these peaks indicate that the molecules are tilted with respect to the bilayer normal and that the direction of the tilt is toward nearest neighbor [50].

#### A. Materials and methods

1,2-dipalmitoyl-sn-glycero-3-phosphocholine (DPPC) and 1,2-dimyristoyl-sn-glycero-3-phosphocholine (DMPC) were purchased from Avanti Polar, and were used as received. Aligned multibilayers of binary lipid mixtures were prepared as follows: A concentrated solution of DPPC-DMPC binary mixtures [dissolved in a mixture of chloroform and methanol

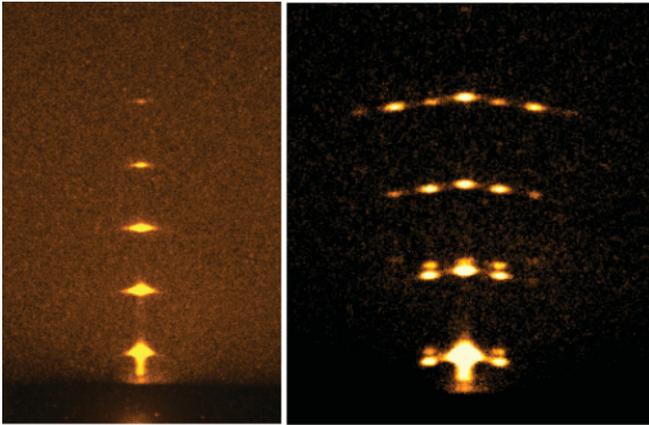


FIG. 8. (Color online) Small-angle diffraction patterns of aligned samples of DPPC-DMPC mixtures in the  $L_{\alpha}$  phase at  $T = 45.0^{\circ}\text{C}$ ,  $\text{RH} = 99.9\%$ ,  $\phi = 0.5$  (left),  $P_{\beta'}$  phase at  $T = 30.1^{\circ}\text{C}$ ,  $\text{RH} = 99.9\%$ ,  $\phi = 0.5$  (right). The bilayer normal is vertical.

(1:1)] in designated molar ratios was deposited on the outer surface of a clean cylindrical glass substrate (radius of curvature of 9 mm). After deposition, the samples were placed overnight under vacuum to remove any remaining traces of the solvent. Subsequently, they were kept in a water-saturated atmosphere and were hydrated for a couple of days to obtain a stack of bilayers oriented parallel to the surface.

Cu  $K_{\alpha}$  ( $\lambda = 1.54 \text{ \AA}$ ) radiation from a rotating anode x-ray generator (Rigaku, Ultra X18) operating at 48 kV and 70 mA and rendered monochromatic by a multilayer mirror (Xenocs) was used to illuminate the hydrated sample kept inside a sealed chamber with two mylar windows. The chamber temperature was controlled using a circulating water bath to an accuracy of  $\pm 0.1^{\circ}\text{C}$  and the relative humidity (RH) inside it was maintained at  $98 \pm 2\%$ , by keeping a reservoir of water. The axis of the cylindrical substrate was oriented perpendicular to the incoming x-ray beam, such that the beam is incident tangentially to the sample.

Diffraction patterns were recorded on a 2D image plate detector of 235 mm diameter and 0.1 mm pixel size (Marresearch). All the samples were first heated to a temper-

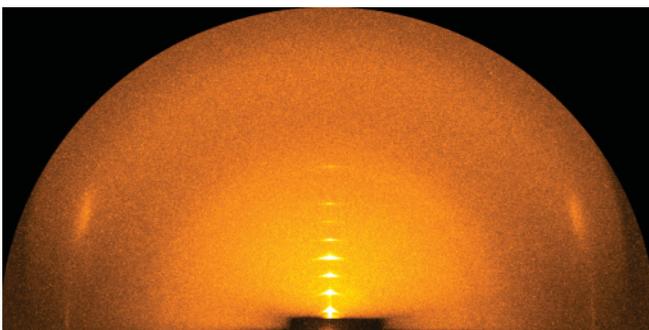


FIG. 9. (Color online) Diffraction pattern of an aligned sample of DPPC-DMPC mixtures in the  $L_{\beta'}$  phases, showing both the small-angle lamellar peaks due to the periodic stacking of the bilayers and the wide-angle peaks due to the in-plane ordering of the tilted chains in the bilayer.  $T = 14.9^{\circ}\text{C}$  and  $\text{RH} = 99.9\%$ ,  $\phi = 0.5$ . The bilayer normal is vertical.

ature above the main transition temperature of DPPC and the diffraction patterns were recorded during cooling from the  $L_{\alpha}$  phase. The sample temperature and the RH close to the sample were measured with a thermohygrometer (Testo 610) inserted into the chamber.

## B. Results and discussion

Figure 10 shows the partial phase diagram of DPPC-DMPC mixtures at  $98 \pm 2\%$  RH. The three pure phases extend over the complete composition range, with two two-phase regions separating them. The phases were identified from their diffraction patterns. The  $L_{\alpha}$  phase gives a few reflections in the small-angle regions, which correspond to different diffraction orders from a periodic lamellar stack. The lamellar periodicity in this phase is  $\sim 5.6 \text{ nm}$  for DPPC and  $\sim 5.0 \text{ nm}$  for DMPC; the mixtures have intermediate values depending on the composition. The wide-angle reflection due to chain ordering is very diffuse in this phase and cannot be seen in the present geometry for exposure times of the order of 10 min. Additional satellite reflections appear in the  $P_{\beta'}$  phase due to the periodic height modulations of the bilayer. These reflections can be indexed on a two-dimensional oblique lattice. The wavelength of the bilayer height modulations in this phase is typically  $\sim 150 \text{ nm}$ . The lower temperature  $L_{\beta'}$  phase shows more lamellar reflections in the small-angle region compared to the  $L_{\alpha}$  phase due to decreased thermal undulations of the rigid bilayers. In addition sharp wide-angle reflections are observed in this phase due to the ordering of the chains in the plane of the bilayer. The presence of an on-axis ( $q_z = 0$ ) and an off-axis ( $q_z \neq 0$ ) reflection show that the chains are tilted, with the tilt direction along the nearest neighbor. The DPPC-DMPC phase diagram closely resembles that calculated from our model for low values of  $J$ . The low value of  $J$  in this case is understandable since the two lipids are very similar with the only difference being that each chain of DPPC is longer by two  $\text{CH}_3$  groups.

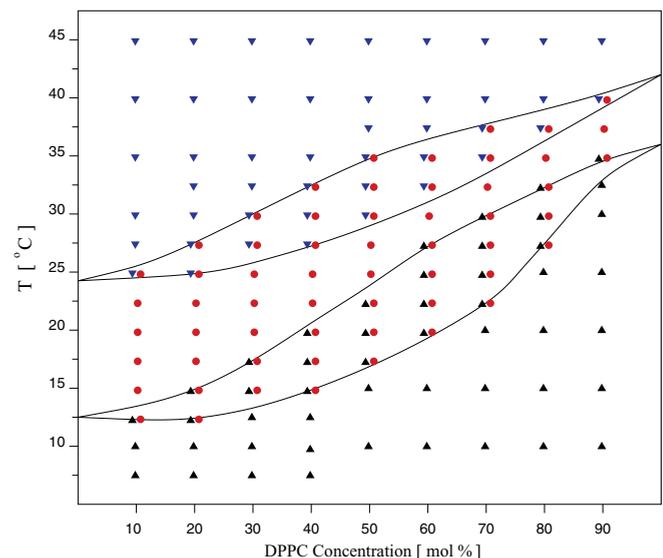


FIG. 10. (Color online) Phase diagram of DPPC-DMPC bilayers at 98% RH determined from x-ray diffraction data.  $\nabla$ ,  $\bullet$ , and  $\triangle$  correspond to the  $L_{\alpha}$ ,  $P_{\beta'}$ , and  $L_{\beta'}$  phases, respectively.

## V. CONCLUSION

We have developed a continuum theory of phase transitions in single- and two-component achiral lipid bilayers. This model is found to reproduce all the salient structural features of the ripple phase occurring between the fluid  $L_\alpha$  phase and the gel  $L_{\beta'}$  phase. In addition, it predicts the existence of a variant

of this phase, which has not been experimentally observed so far. This theory has been used to calculate some generic phase diagrams of binary phospholipid mixtures. We have also determined the phase diagram of DPPC-DMPC mixtures from x-ray diffraction studies on aligned multilayers. It is found to be in good agreement with that calculated from the model for weakly segregating species.

- 
- [1] A. Tardieu, V. Luzzati, and F. C. Reman, *J. Mol. Biol.* **75**, 711 (1973).
- [2] T. J. MacIntosh, *Biophys. J.* **294**, 237 (1980).
- [3] M. Kodama and T. Miyata, *Colloids Surf. A* **109**, 283 (1996).
- [4] M. J. Janiak, D. M. Small, and G. G. Shipley, *J. Mol. Biol.* **254**, 6068 (1979).
- [5] G. S. Smith, E. B. Sirota, C. R. Safinya, and N. A. Clark, *Phys. Rev. Lett.* **60**, 813 (1988);
- [6] G. S. Smith, E. B. Sirota, C. R. Safinya, R. J. Plano, and N. A. Clark, *J. Chem. Phys.* **92**, 4519 (1990)
- [7] J. F. Nagle, *Annu. Rev. Phys. Chem.* **31**, 157 (1980); J. F. Nagle and D. A. Wilkinson, *Biophys. J.* **23**, 159 (1978)
- [8] E. J. Luna and H. M. McConnell, *Biochim. Biophys. Acta* **470**, 303 (1977).
- [9] J. A. N. Zasadzinski, J. Scheir, J. Gurley, V. Elings, and P. K. Hansma, *Science* **239**, 1013 (1988).
- [10] M. P. Hentschel and F. Rustichelli, *Phys. Rev. Lett.* **66**, 903 (1991).
- [11] D. C. Wack and W. W. Webb, *Phys. Rev. Lett.* **61**, 1210 (1988).
- [12] K. Mortensen, W. Pfeiffer, E. Sackmann, and W. Knoll, *Biochim. Biophys. Acta* **945**, 221 (1988).
- [13] R. Krbecek, C. Gebhardt, H. Gruler, and E. Sackmann, *Biochim. Biophys. Acta* **554**, 1 (1979).
- [14] A. Hicks, M. Dinda, and M. A. Singer, *Biochim. Biophys. Acta* **903**, 177 (1987).
- [15] J. A. N. Zasadzinski, *Biochim. Biophys. Acta* **946**, 235 (1988).
- [16] H. W. Meyer, *Biochim. Biophys. Acta* **1302**, 138 (1996).
- [17] J. T. Woodward and J. A. Zasadzinski, *Biophys. J.* **72**, 964 (1997).
- [18] W.-J. Sun, S. Tristram-Nagle, R. M. Suter, and J. F. Nagle, *Proc. Natl. Acad. Sci.* **93**, 7008 (1996).
- [19] W.-J. Sun, R. M. Suter, M. A. Knewton, C. R. Worthington, S. Tristram-Nagle, R. Zhang, and J. F. Nagle, *Phys. Rev. E* **49**, 4665 (1994).
- [20] K. Sengupta, V. A. Raghunathan, and J. Katsaras, *Phys. Rev. E* **59**, 2455 (1999).
- [21] K. Sengupta, V. A. Raghunathan, and Y. Hatwalne, *Phys. Rev. Lett.* **87**, 055705 (2001).
- [22] K. Sengupta, V. A. Raghunathan, and J. Katsaras, *Europhys. Lett.* **49**, 722 (2000).
- [23] S. Doniach, *J. Chem. Phys.* **70**, 4587 (1979).
- [24] M. S. Falkovitz, M. Seul, H. L. Frisch, and H. M. McConnell, *Proc. Natl. Acad. Sci.* **79**, 3918 (1982).
- [25] M. Marder, H. L. Frisch, J. S. Langer, and H. M. McConnell, *Proc. Natl. Acad. Sci.* **81**, 6559 (1984).
- [26] J. M. Carlson and J. P. Sethna, *Phys. Rev. A* **36**, 3359 (1987).
- [27] R. E. Goldstein and S. Leibler, *Phys. Rev. Lett.* **61**, 2213 (1988).
- [28] R. E. Goldstein and S. Leibler, *Phys. Rev. A* **40**, 1025 (1989).
- [29] T. C. Lubensky and F. C. MacKintosh, *Phys. Rev. Lett.* **71**, 1565 (1993).
- [30] C.-M. Chen, T. C. Lubensky, and F. C. MacKintosh, *Phys. Rev. E* **51**, 504 (1995).
- [31] K. Honda and H. Kimura, *J. Phys. Soc. Jpn.* **60**, 1212 (1991).
- [32] U. Seifert, J. Shillcock, and P. Nelson, *Phys. Rev. Lett.* **77**, 5237 (1996).
- [33] J. Katsaras, R. F. Epanand, and R. M. Epanand, *Phys. Rev. E* **55**, 3751 (1997).
- [34] J. Katsaras and V. A. Raghunathan, *Phys. Rev. Lett.* **74**, 2022 (1995).
- [35] M. A. Kamal, A. Pal, V. A. Raghunathan, and M. Rao, *Europhys. Lett.* **95**, 48004 (2011).
- [36] R. P. Rand, *Annu. Rev. Biophys. Bioeng.* **10**, 277 (1981).
- [37] N. Shimokawa, S. Komura, and D. Andelman, *Eur. J. Phys. E* **26**, 197 (2008).
- [38] A. E. Jacobs, C. Grein, and F. Marsiglio, *Phys. Rev. B* **29**, 4179 (1984).
- [39] L. Yang and M. Fukuto, *Phys. Rev. E* **72**, 010901 (2005).
- [40] K. Sengupta, V. A. Raghunathan, and J. Katsaras, *Phys. Rev. E* **68**, 031710 (2003).
- [41] R. J. Wittebort, C. F. Schmidt, and R. G. Griffin, *Biochemistry* **20**, 4223 (1981).
- [42] M. B. Schneider, W. K. Chan, and W. W. Webb, *Biophys. J.* **43**, 157 (1983).
- [43] A. H. de Vries, S. Yefimov, A. E. Mark, and S. J. Marrink, *Proc. Natl. Acad. Sci.* **102**, 5392 (2005).
- [44] O. Lenz and F. Schmid, *Phys. Rev. Lett.* **98**, 058104 (2007).
- [45] D. Jamróz, M. Kepczynski, and M. Nowakowska, *Langmuir* **26**, 15076 (2010).
- [46] M. Kranenburg and B. Smit, *J. Phys. Chem. B* **109**, 6553 (2005).
- [47] K. Simons and J. L. Sampaio, *Cold Spring Harb. Perspect. Biol.* **3**, a004697 (2011).
- [48] A. Blume, R. J. Wittebort, S. K. Das Gupta, and R. G. Griffin, *Biochemistry* **21**, 6243 (1982).
- [49] S. H. Wu and H. M. McConnell, *Biochemistry* **14**, 847 (1975).
- [50] M. Hentschel and R. Hosemann, *Mol. Cryst. Liq. Cryst.* **94**, 291 (1983).